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Edited by
Phillipe Collery
Peter Brätter
Virginia Negretti de Brätter
Lylia Khassanova
Jean-Claude Etienne



## Absorption of silicon in healthy subjects

M.R. Calomme<sup>1</sup>, P. Cos<sup>1</sup>, P.C. D'Haese<sup>2</sup>, R. Vingerhoets<sup>1</sup>, L.V. Lamberts<sup>2</sup>, M.E. De Broe<sup>2</sup>, C. Van Hoorebeke<sup>1</sup>, D.A. Vanden Berghe<sup>1</sup>

<sup>1</sup> Department of Pharmaceutical Sciences; <sup>2</sup> Department of Nephrology-Hypertension, University of Antwerp (UIA), Universiteitsplein 1, B-2610 Antwerp, Belgium

## 1. INTRODUCTION

Silicon (Si) is next to oxygen the most abundant element in the earth's crust. The distribution of Si in vertebral tissue and physiological changes in bone caused by dietary Si deficiency indicate that Si influences bone formation by affecting cartilage composition and ultimately cartilage calcification [1]. The dietary intake of Si was estimated for US citizens to be between 20-50 mg with the lower intake for animal-based diets and the higher intake for plant-based diets [2]. However, studies on the minimum Si requirement and supplementation experiments, comparing the bioavailability of different Si compounds, are lacking. Orthosilicic acid was suggested to have an important function in Si metabolism [1, 6] and is found in both fresh water and sea water. This monomeric form of silicic acid is stable in dilute concentrations of about 10-4 M, but condenses into silica gels at higher concentrations and low pH. The absorption and urinary excretion of Si from (a) stabilized orthosilicic acid (OSA), (b) standardized herbal silica extract from the Si-accumulator plant Equisetum arvense, and (c) colloidal silicic acid were compared in a double-blind study with healthy subjects.

## 2. MATERIALS AND METHODS

Fourteen healthy subjects (8 females, 6 males, aged 22-34 y) were included after informed, written consent. None had taken Si supplements within 3 months before the start of the study. Each fasting subject received in a cross-over protocol an equimolar dose of Si p.o. (20 mg Si) in the form of stabilized orthosilicic acid (OSA, 1ml of BioSil containing 20 g Si/l as stabilized monomeric silicic acid, Bio Minerals NV, Belgium [3]), herbal silica (533 mg of a dry Equisetum arvense extract containing 8 % (w/w) SiO<sub>2</sub>), colloidal silicic acid (2 ml of 28 g H<sub>2</sub>SiO<sub>3</sub>/l), or a placebo (10 ml mineral water) with 1 week wash-out period between each supplement or the placebo. Blood samples were collected in Si free polypropylene tubes prior to supplementation and after 1, 2, 4, 6, and 8 hours post partem. Urine was collected between 0 and 24 hours post partem in polypropylene containers. Identical meals were consumed during the experiment after 2

and 6 hours supplementation. The Si concentration in serum and urine was determined for each subject in one batch with AAS [4]. A Zeeman/3030 Atomic Absorption Spectrometer equipped with a HGA-600 graphite furnace was used in combination with an AS-60 autosampler (Perkin-Elmer Corp. Norwalk CT). Pyrolytic coated graphite tubes were used. The hollow cathode lamp settings were respectively 30 mA lamp current, 251.6 nm spectral line and 0.7 nm band width. The injected sample volume was 10 µl and signals were measured in the peak-area mode. The furnace program is shown in table I. Serum and urine samples were diluted 1/10 in a matrix-modifier solution containing 0.125 g/l La(NO<sub>3</sub>)<sub>3</sub>.6H2O; 0.111 g/l CaCl<sub>2</sub>; 1.50 g/l NH<sub>4</sub>H<sub>2</sub>PO<sub>4</sub>; 0.50 g/l Na<sub>4</sub>EDTA.4H<sub>2</sub>O and 0.1 % (v/v) HNO<sub>3</sub>. Samples were measured against a calibration curve prepared in the matrix-modifier solution, using as standards 0 µg/l, 62.5 µg/l, 125 µg/l and 250 µg/l Si, respectively. The sensitivity determined as the amount of silicon yielding a 0.0044 Abs.s signal was 82 pg. The precision determined as the inter-assay c.v. was 13.3%, 6.2%, and 3.1% for Si concentrations of 240 µg/l (n=6), 627 µg/l (n=6), and 1823 µg/l (n=6), respectively.

The area under the time concentration curve (A.U.C.) was calculated using the linear trapezoidal rule. Statistical significance was investigated with the Wilcoxon matched-pairs signed ranked-test. The relation between two parameters was investigated with the Pearson correlation procedure.

#### 3. RESULTS

The mean baseline silicon concentration in serum was lower compared to reported values for serum of normal subjects (table II).

A significant increase in serum Si concentration (fig. 1) from the baseline value was observed after respectively 1 h for OSA (mean  $\pm$  S.E.:  $54 \pm 19 \,\mu\text{g/l}$ , p<0.005), 4 h for the placebo (23  $\pm$  11  $\,\mu\text{g/l}$ , p<0.025) and herbal silica (37  $\pm$  14  $\,\mu\text{g/l}$ , p<0.025), and 8 h for colloidal Si (36  $\pm$  12  $\,\mu\text{g/l}$ , p = 0.01). The mean A.U.C. (table III) was significantly higher after OSA supplementation (p<0.005), but was not significantly different for respectively herbal silica and colloidal Si compared to the placebo.

A significant correlation was found between the individual A.U.C. and the individual urinary Si concentration (r = 0.28, p < 0.05, n = 56). The urinary silicon excretion (table 2) was significantly higher after OSA supplementation (p < 0.005) but was not significantly different for respectively herbal silica and colloidal Si compared to the placebo.

## 4. DISCUSSION

Most likely, the fasting condition of the subjects in the present study explains the lower Si concentration in serum compared to values reported in the literature. In fact, the serum Si concentration of subjects receiving a placebo increased 2 hours after consumption of a meal, indicating a significant variation of the serum level during the day.

Table I
Temperature programming of furnace for the analysis of Si in serum and urine.

Temperature (°C)	Ramp time (s)	Hold time (s)	Gasflow <sup>a</sup> (ml/min.)
100	15	15	300
120	10	10	300
700	5	5	300
1400	60	10	300
2500	0	4	0
2700	1	3	300

<sup>&</sup>lt;sup>a</sup>Argon was used as purge gas.

Table II
Silicon concentration in serum of fasting, healthy subjects compared to serum concentrations reported in the literature for normal adults.

Author, (year of publication)	Number (female, male)	Method	Si conc. (μg/l)
Dobbie & Smith (1982), [5]	50 (25, 25)	AAS	$603 \pm 126^{b}$
Berlyne et al. (1986), [6]	23 (2, 21)	AAS	$265 \pm 17^{\circ}$
Hosokawa et al. (1990), [7]		AAS	$200 \pm 60^{b}$
Bercowy et al. (1994), [8]	365 (- <sup>a</sup> )	DCP-AES	<200 - 680 <sup>d</sup>
Teuber et al. (1995), [9]	55 (55, 0)	ICP-AES	$130 \pm 70^{b}$
D'Haese et al. (1995), [10]	_•	AAS	$161 \pm 44^{b}$
Calomme et al., (1998),	14 (8, 6)	AAS	$112 \pm 42^{b}$
[present study]			<u></u>

<sup>\*</sup>Not specified; bmean ± S.D.; cmean ± S.E.; drange.

Table III
Silicon absorption and excretion from different silicon sources in healthy subjects.

Group	Number	Si absorption 0 - 8 hours p.p. A.U.C. (µgh/l)	Urinary Si excretion (μg/g creatinine) <sup>a</sup>
Placebo	14	202 ± 37	$13.3 \pm 1.5$
Stabilized orthosilicic acid (OSA)	14	659 ± 95 <sup>b</sup>	$17.5 \pm 0.9^{b}$
Herbal silica	14	$248 \pm 53^{\circ}$	$12.0 \pm 1.1^{\circ}$
Colloidal silicic acid	14	164 ± 47°	$13.8 \pm 2.1^{\circ}$

<sup>\*</sup>Mean ± S.E.; bp<0.002 versus placebo; cnot significantly different versus placebo.

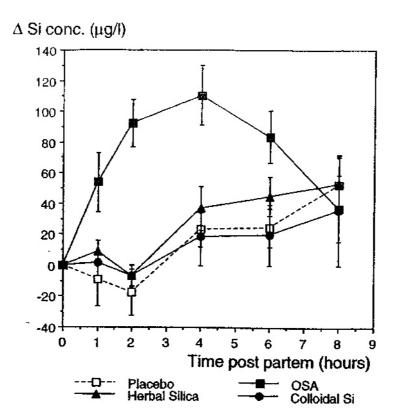


Fig. 1 Increase in serum Si concentration from the baseline value in healthy subjects. An equimolar dose of Si (20 mg) was given in the form of stabilized orthosilicic acid (OSA), herbal silica, and colloidal silicic acid. Mean values  $\pm$  S.E. are shown (n = 14)

The present study demonstrates that the bioavailability of silicon is largely dependent on its chemical form. The absorption of Si is faster, higher and less subject dependent for stabilized orthosilicic acid compared to herbal silica and colloidal silicon which are polymerized forms of orthosilicic acid. Consequently, the chemical form of Si in the diet will influence the minimal Si requirement for humans.

The difference in absorption between Si supplements is confirmed by a higher urinary Si excretion for OSA and is in agreement with the dose-dependent Si absorption from stabilized orthosilicic acid which was demonstrated in an animal supplementation study [3]. The significant correlation between the absorption measured by the A.U.C. and the urinary excretion confirms earlier reports suggesting renal clearance as a major route for silicon excretion.

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